



## TOXICOPATHOLOGICAL AND IMMUNOTOXICITY INDUCE BY CYGON IN GASTROINTESTINAL TRACT OF ALBINO MICE

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Article Received date: 04 April 2025

Article Revised date: 26 April 2025

Article Accepted date: 15 May 2025



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### ABSTRACT

The purpose of the current study was to examine the harmful effects of cygon in mice, as well as the impact of immunological stimulation with Aftovaxpur DOE vaccine and histopathological abnormalities. Eighty-four-week-old adult Swiss Albino mice were split up into four groups. The first group received 4.5 mg/kg of Cygon orally every day for four weeks. The second group was given 0.1 ml of the Aftovaxpur DOE vaccine intraperitoneally (I/P) twice at two-week intervals while also receiving cygon orally every day for four weeks. I/P administered 0.1 ml of the Aftovaxpur DOE vaccine to the third group. The negative control was assigned to the fourth group. Samples of blood were drawn from the first, second, third, and fourth groups in order to measure serum IL8, IL10 levels using an ELISA kit. The results of ELISA tests for detecting IL8 and IL10 titers in group three ( $P \leq 0.05$ ) demonstrated a substantial increase ( $P \leq 0.05$ ) when compared to group two, group one, and group four. Clinical signs of treated mice showed depression and decreased appetite throughout the trial period, the anorexia and weight loss were brought on by pathological lesions that resulted in intestinal malabsorption and stomach indigestion. Pathological lesions of the intestine showed desquamation in epithelial cells and mucus accumulation in addition to necrotizing enteritis also degeneration of mucous glands and inflammatory cells infiltration between mucous glands, as well as hyperplasia of goblet cells, in stomach congestion of blood vessels and moderate hyperkeratosis in epithelial cells. Even so, the groups that received Cygon treatment and the Aftovaxpur DOE vaccination showed improvement against the negative effects of Cygon.

**KEYWORDS:** cygon Histopathological changes, Immunotoxicity, intestine, Cygon, IL8, IL10

### INTRODUCTION

Cygon is a common organophosphate pesticide and acaricide. American Cyanamid patented and marketed it in the 1950s. Cygon, like other organophosphates, is an acetylcholinesterase inhibitor, meaning it inhibits cholinesterase, an enzyme required for central nervous system function. It works through both touch and ingestion. It is easily absorbed and disseminated throughout tissues, and it degrades quickly (Hulse *et al.*, 2019). Cygon is a organophosphate that is insoluble in alcohol and acetone but soluble in water. It also has a high lipophilicity. Crystals that are color-less, white, or light beige can be found (WHO).

Few studies on the histopathological effects of Cygon (Persis, 2001) in animals have been conducted. Furthermore, Cygon causes hyperglycemia and a variety of harmful effects on the rat pancreas after acute, subchronic, and chronic exposure (Kamath *et al.*, 2008). In experimental animals, pesticides have harmful effects.

It has the ability to affect biochemical, hormonal, reproductive, and oxidative stress indicators. In experimental animals, it also has cytotoxic, autogenetic, genotoxic, and carcinogenic properties (Mossa *et al.*, 2017). Oxidative stress, and the subsequent destruction to critical cell components induced by oxygen-free radicals, is often regarded as a significant process (Livingstone, 2001).

Hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>)-induced oxidative stress is known to cause a wide spectrum of physiological, biochemical, and immunological dysfunctions in experimental animals and people. Furthermore, it triggers cell death and induces apoptosis as a result of DNA damage (Ramond *et al.*, 2011; Lumaet *et al.*, 2013)

Previous research suggested that some pesticide-related effects could be caused by changes in membrane fluidity (Mansour and Mossa, 2009), lipid composition and inhibition of enzyme activities (Mansour and Mossa,

2010). IL-8 was initially identified as a neutrophil-specific chemotactic factor before being categorized as, a member of the chemokine family. Following interleukin-1a, 1b, 17, tumor necrotic factor alpha, or TLR activation, a variety of cells, including macrophages, neutrophils, lymphocytes, and endothelial and epithelial cells, produce IL-8 (Coelho *et al* 2005). The activation of neutrophils and their migration to the site of infection or injury are the main effector activities of IL-8. In addition to neutrophils, NK cells, T cells, basophils, and eosinophils primed with GM-CSF or IL-3 are also drawn to IL-8. (Burke 2008).

Huda (2020) showed that in convalescent patients, the modulation of innate immune response coupled with the establishment of adaptive immunity neutralizing antibodies, memory T and B cells provides optimism for active immunization. Examining the immunotoxic and histopathological effects of Cygon on the digestive system was the aim of this study.

## MATERIAL AND METHODS

### Chemicals

Cygon, is an organophosphorous pesticide with achemical formula:  $\text{SCH}_3\text{NHCOCH}_2\text{SP}(\text{OCH}_3)_2$

**Trade name:** cekuthoatedimetferkethion, roxion, rego, afidox, rogodial, rogodan, trimetion and sevigordimethoate as moderatelyhazardous.

Preparation of cygon solution:

To prepare the cygon solution the following procedure was followed. About 90 milligrams of cygon(powder) was dissolved in 200 ml of distilled water and the final concentration was 90 mg/200ml. For each 10 g of body

weight of mice, 0.1 mL of cygon solution was administrated orally to reach 4.5 mg/kg.BW.(Sabina 2001).

### Aftovaxpur DOE preparation

Animals were immunized with this antigen made in accordance with House *etal.* (1989).

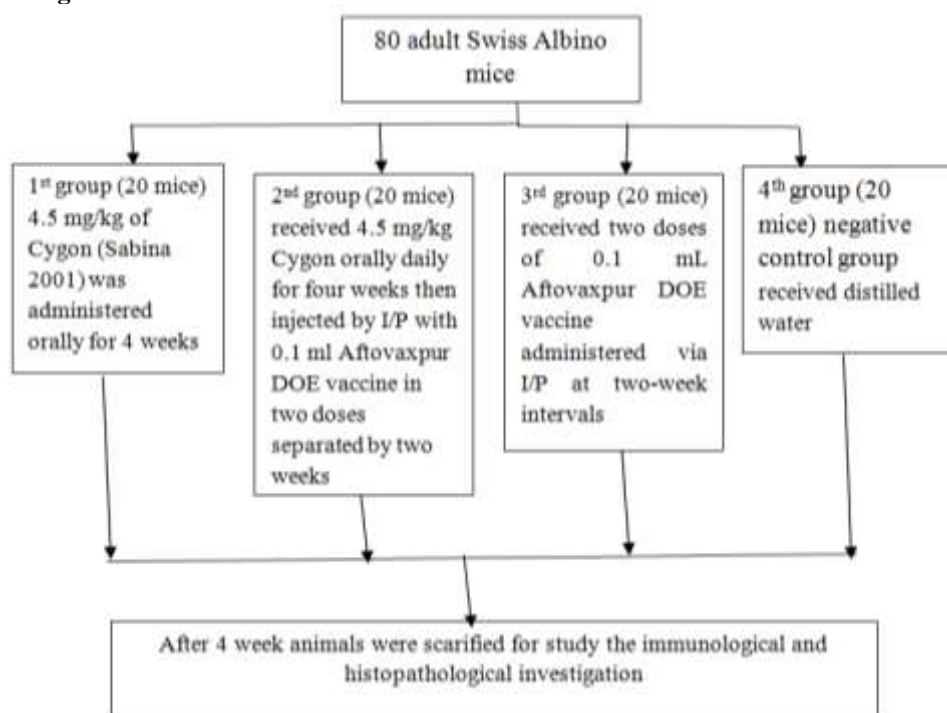
### Experimental Animals

For adaptation, 80 adult Swiss Albino mice weighing between 25 and 30 grams at 4 weeks of age were acquired from the animal house of the College Vet. Med. University of Baghdad. The animals were kept in plastic cages in a room with air conditioning that was kept at  $25\pm 2^\circ\text{C}$ . The bedding in the cages was made of hard wood chips, and it was changed frequently to maintain a hygienic environment. Mice were provided with food pellets and unlimited water.

### Immunological tests

Blood was drawn directly from the mice hearts; for **Immunological** testing, placed in coagulant test tube, blood samples were centrifuged for five minutes at 3000 rounds per minute (rpm), and serum was extracted. It was then frozen at  $-20^\circ\text{C}$  until it was needed for **Immunological** test (serum level of IL8, IL10). An enzyme-linked immunosorbent assay (ELISA) kit from Elabscience (U.S.A.) was used to measure the concentrations of IL8 and IL10 in mouse serum, this sandwich kit is for the accurate quantitative detection of Mouse Interleukin 8,10(also known asIL-8, IL10) in serum, plasma, cell culture supernates, Ascites, tissue homogenates or other biological fluids. this test was carried out in accordance with the manufacturer's procedure.

### Experimental Design



### Histopathological examination

At the conclusion of the four-week experiment, all animals were killed, and stomach and intestine samples were taken. After fixing the tissues in a 10% formaldehyde solution, we used a histokite to process them as usual. Hematoxylin and eosin staining and microtome were used to section paraffin blocks containing implanted tissue slices, which were subsequently examined under a light microscope (Luna, 1968).

### Statistical analysis

Statistical analysis was conducted using a two-way ANOVA, and the statistical package for social sciences

(SPSS) revealed that the mean difference was significant at the ( $P \leq 0.05$ ) level.

## RESULTS

### ELISA test for detection levels of IL8 and IL10 titer

The final result of ELISA test showed that the mean values of serum IL8 and IL10 (mg/dl), at 28 day Group 3's post-immunization rate was noticeably significantly increase ( $266.84 \pm 1.04$ ,  $143.30 \pm 0.20$ ), then those values in the G1, G2 ( $5.80 \pm 1.12$ ,  $7.71 \pm 0.09$ ), ( $44.55 \pm 1.05$ ,  $58.00 \pm 0.22$ ) and negative control groups G4 ( $11.45 \pm 0.13$ ,  $8.45 \pm 1.01$ ) respectively, as shown in Table 1.

**Table (1): The mean value of serum IL8 and IL10 in immunized groups and control negative group at 28 day post immunization by ELISA test.**

Groups	IL8 (mg/dl) (Mean $\pm$ SE) at 4 weeks	IL10 (mg/dl) (Mean $\pm$ SE) at 4 weeks
G1	$5.80 \pm 1.12$ D	$7.71 \pm 0.09$ D
G2	$44.55 \pm 1.05$ B	$58.00 \pm 0.22$ B
G3	$266.84 \pm 1.04$ A	$143.30 \pm 0.20$ A
G4	$11.45 \pm 0.13$ C	$8.45 \pm 1.01$ C

Means with distinct capital letters differ significantly in the same column are significantly differ ( $P \leq 0.05$ ).

### Clinical signs of treated mice

All of the treated mice showed signs of depression and decreased appetite throughout the trial period. The anorexia and weight loss were brought on by pathological lesions that resulted in intestinal malabsorption and stomach indigestion, whereas the control group consumed normally.

### Histopathological Examination

#### The histopathological changes in mice of (1<sup>st</sup> group) Intestine

The microscopic section of the intestine showed desquamation in epithelial cells and mucus accumulation (Figure 1) in addition to necrotizing enteritis (Figure 2), also degeneration of mucous glands and inflammatory cells infiltration between mucous glands (Figure 3), as well as hyperplasia of goblet cells (Figure 4).

### Stomach

The microscopic section of the stomach showed congestion of blood vessels (Figure 5), as well as moderate hyperkeratosis in epithelial cells (Figure 6).

#### The histopathological changes in mice of 2<sup>nd</sup> group Intestine

The microscopic section of the intestine showed necrosis in epithelial cells and inflammatory cells infiltration in lamina propria (Figure 7), in other section showed mucus accumulation in lumen (Figure 8).

#### The histopathological changes in mice of 3<sup>rd</sup> group Intestine

The microscopic section of the intestine showed hyperplasia of lymphoid tissue (Figure 9) in other section showed proliferation of goblet cells in lamina propria (Figure 10).



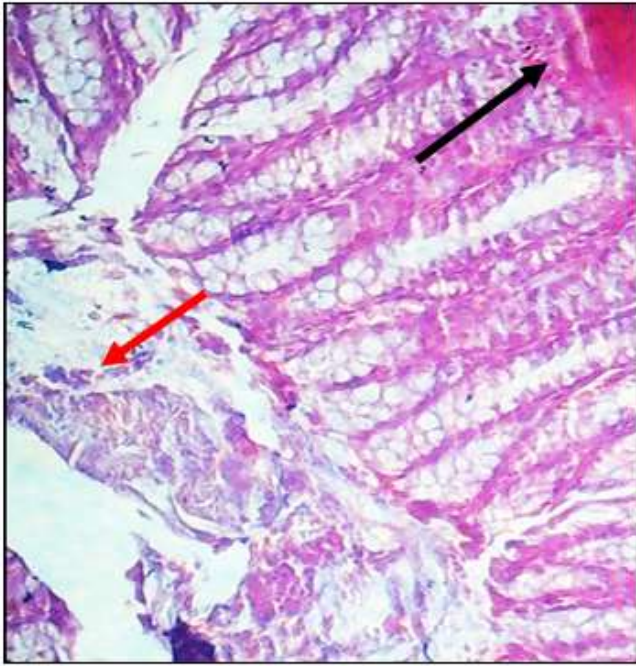


Figure (1): Histopathological section of intestine at 4 weeks in 1<sup>st</sup> group shows desquamation in epithelial cells (black arrow), mucus accumulation (red arrow) (black arrow) (H and E stain 100X)

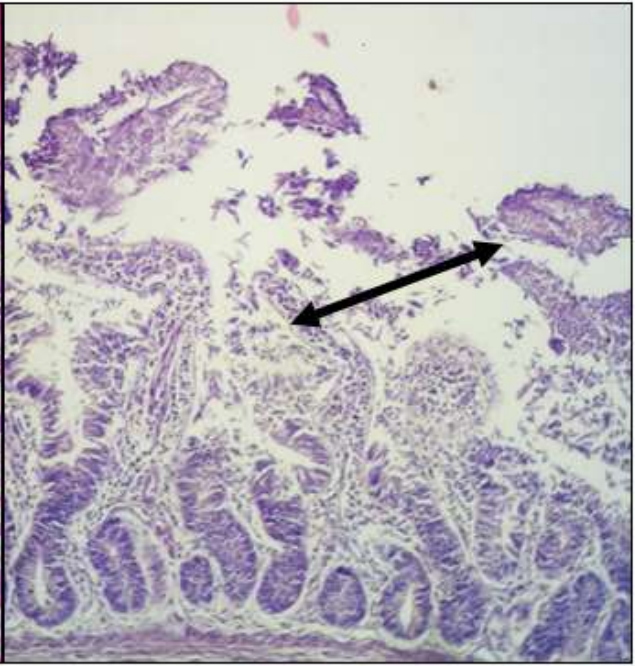


Figure (2): Histopathological section of intestine at 4 weeks in 1<sup>st</sup> group shows necrotizing enteritis (black arrow) (H and E stain 100X)

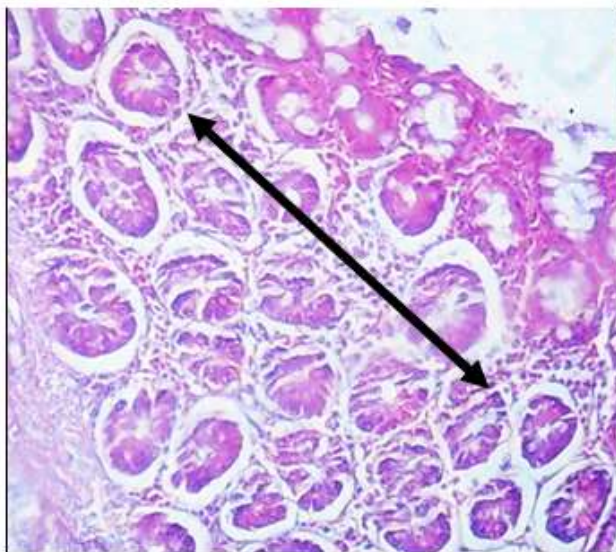


Figure (3): Histopathological section of intestine at 4 weeks in 1<sup>st</sup> group show degeneration of mucous glands and inflammatory cells infiltration between mucous glands (black arrow) (H and E stain 400X)

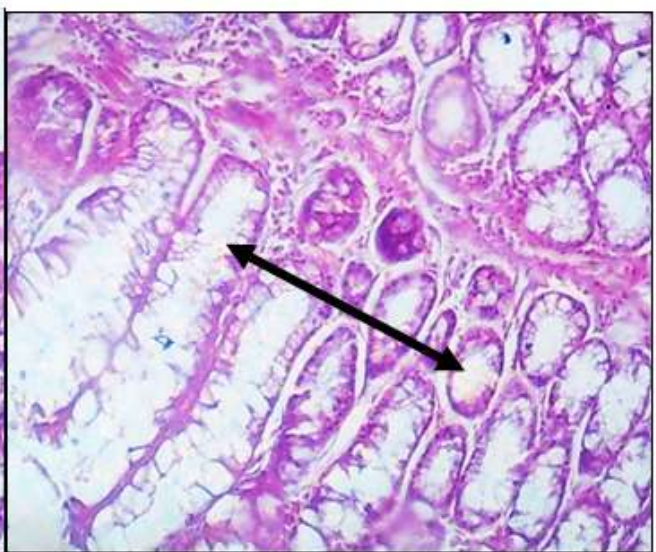


Figure (4): Histopathological section of intestine at 4 weeks in 1<sup>st</sup> group show hyperplasia of goblet cells (black arrow) (H and E stain 400X)



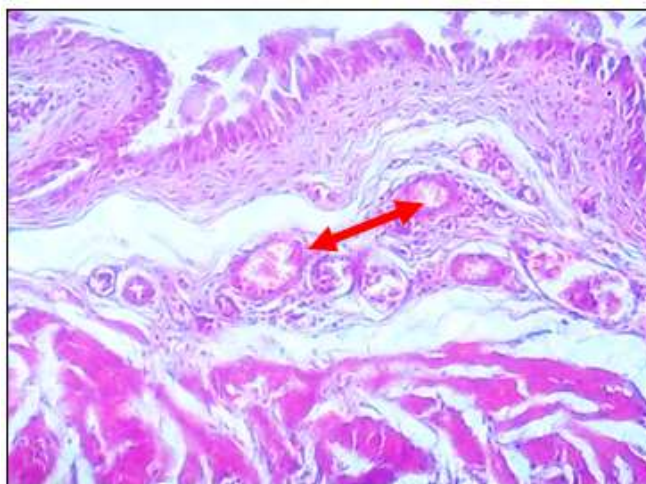


Figure (5): Histopathological section of stomach at 4 weeks in 1<sup>st</sup> group shows, congestion of blood vessels (red arrow) (H and E stain 100X)

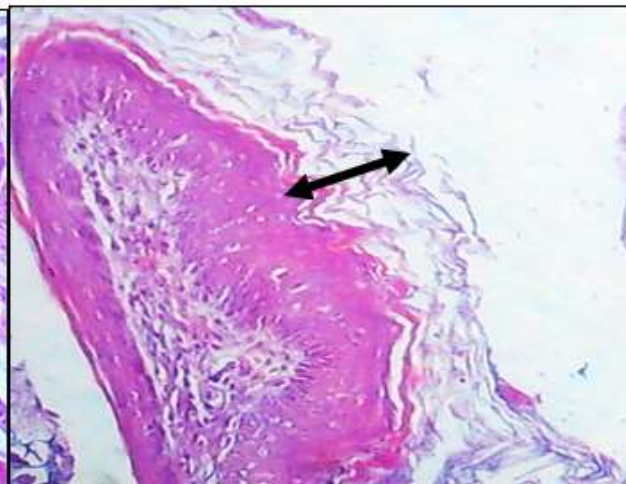


Figure (6): Histopathological section of stomach at 4 weeks in 1<sup>st</sup> group shows, moderate hyperkeratosis in epithelial cells (black arrow) (H&E stain, 100×).

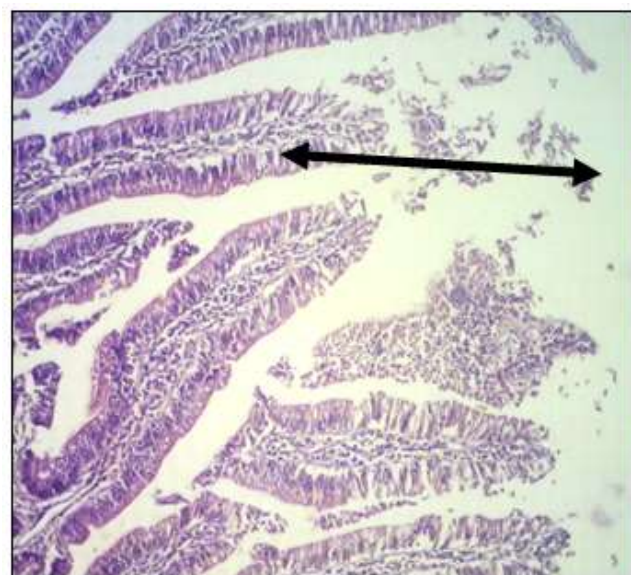


Figure (7): Histopathological section of intestine at 4 weeks in 2<sup>nd</sup> group shows necrosis in epithelial cells and inflammatory cells infiltration in lamina propria) (black arrow) (H&E stain, 100×).

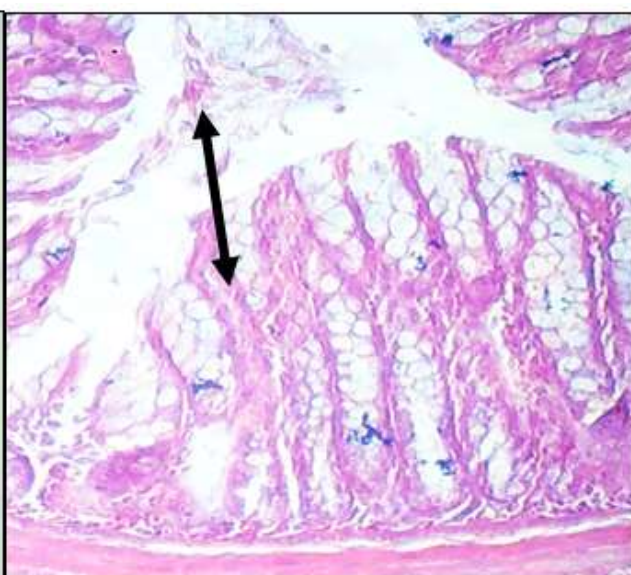


Figure (8): Histopathological section of intestine at 4 weeks in 2<sup>nd</sup> group shows mucus accumulation in lumen (black arrow) (H and E stain 100X)

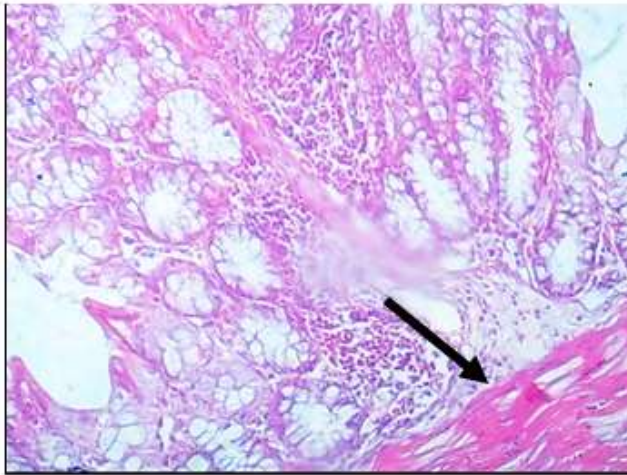


Figure (9): Histopathological section of intestine at 4 weeks in 3<sup>rd</sup> group shows hyperplasia of lymphoid tissue (black arrow) (H&E stain, 100×).

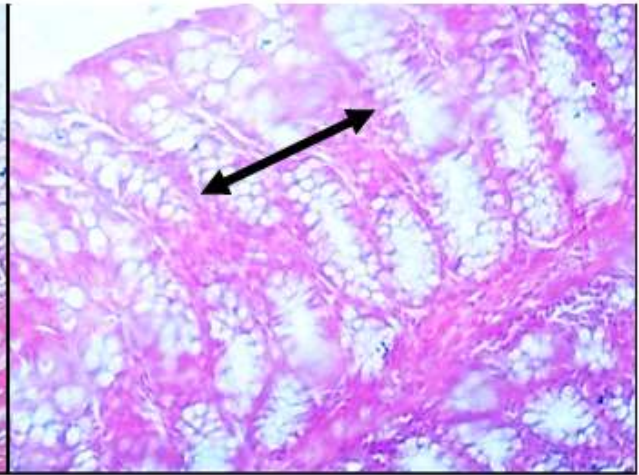


Figure (10): Histopathological section of intestine at 4 weeks in 3<sup>rd</sup> group shows proliferation of goblet cells in lamina propria (black arrow) (H&E stain, 100×).

## DISCUSSION

As previously mentioned, the results may indicate that antigens boosted the immune response while Cygon decreased the Ag's immunological response. These data may also suggest that pesticide contamination in the environment is linked to lower immunization program activity against Aftovaxpur DOE agricultural animals. Furthermore, Cygon lowered immunity and alterations within the suppressor gene, indicating that immunological problems, gene mutations, and oxidative stress can all have an impact on functioning cells. (El Okda *et al.*, 2017) discovered differences in the amounts of the antioxidant-protective enzymes SOD, CAT, GPx, and GSH. Occur during pyrethroid exposure. The immunotoxicity of Cygon was studied in 30 employees (El Okda *et al.*, 2017).

Permethrin also suppressed splenic T-lymphocyte growth. Permethrin inhibited spleen cells growth by inhibiting thymic cell proliferation. Furthermore, higher apoptosis was seen in CD48 and CD48+ thymocytes, although the CD4+8+ thymocyte subpopulation was dramatically decreased (WHO). Permethrin increases oxygen and neutrophil activity. After one hour of incubation with permethrin or its metabolites, neutrophil apoptosis was also observed (Gabbianelli *et al.* 2009). It increases superoxide dismutase and alanine aminotransferase (AAT) in mice (Kumar and Sharma, 2015). By activating caspases, Cygon causes oxidative stress and triggers the mitochondrial death pathway (Kumar and Sasmal, 2018). Caspase-3 activation promotes apoptosis in spleen cells by activating a death protease (Kumar and Sharma, 2015). Cox-2 expression has been investigated for its capacity to induce inflammation, and bcl-2 and p53 proteins have been investigated for their capacity to induce apoptosis (Maalej *et al.*, 2017). Decreases in bcl-2 protein levels and increases in cox-2 and p53 protein levels cause apoptosis.

According to Ma *et al.* (2023), our earlier study demonstrated that prolonged exposure to cygon may cause mitochondrial damage and increased oxidative stress in the colonic epithelial cells of mice, resulting in colonic inflammation.

One of the pathological mechanisms of numerous diseases is oxidative stress (Zheng *et al.*, 2020). Research has indicated that oxidative stress plays a role in the onset of fibrosis (Masarone *et al.*, 2018). The expression of several antioxidant enzymes is regulated by the crucial transcription factor nuclear factor erythroid 2-related factor 2 (Nrf2) (Chen *et al.*, 2020).

A significant inflammatory response will result from the excess ROS (Su *et al.*, 2019). ROS triggers the classic NF- $\kappa$ B inflammatory signaling pathway in pathological conditions. Numerous proinflammatory cytokines, including interleukin-1 $\beta$ , interleukin-6, and TNF- $\alpha$ , are then released, exacerbating the inflammatory damage in the liver (Wei *et al.*, 2018). TNF- $\alpha$  causes tissue damage and persistent inflammation by inducing the production of chemokines by fibroblasts, endothelial cells, and macrophages. Nrf2 is an upstream regulator that controls the oxidative stress response and, by controlling cytokine production, reduces inflammation (Baiyun *et al.*, 2018). Accordingly, the addition of Res decreased DLM-induced inflammation in this trial. Thus, exposure to cygon causes the quail's liver to become inflamed by activating the NF- $\kappa$ B/TNF- $\alpha$  signaling pathway.

On the other hand, the vaccine by itself causes cytokines IL8 and IL10 to increase. According to Mahajan *et al.* (2005), *Brucella melitensis* surface proteins can elicit humoral immune responses as well as cell-mediated immunity. The Th2 response was triggered by spleen cells in mice that were immunized with soluble *Brucella* antigens. The proteins that were secreted by *Brucella* were effective in promoting cell-mediated immunity.



Oxidative stress may be linked to the pathological abnormalities in the examined organs in the group that received Cygon treatment., which is important in the pathophysiology of many intestine and stomach illnesses produced by Cygon therapy. may generate oxidative stress, leading in free radical generation and lipid peroxidation, and may be the underlying molecular mechanism causing pesticide-induced toxicity (Ismail and Mohamed, 2012). By oxidizing lipids, proteins, and DNA, excessive oxidant production can cause oxidative damage in cells. There is emerging evidence that oxidative stress, namely stress caused by reactive oxygen and reactive nitrogen species, can lead to a wide range of inflammatory and degenerative diseases (MacDonald-Wicks *et al.*, 2006). The gut is a critical site of hazardous chemical absorption. Because the gut is thought to be the primary route for pesticide absorption, it is not surprising that the duodenum showed goblet hypertrophy. The pathological changes observed in the intestine and stomach in the current study are consistent with the findings of Velmurugan *et al* (2007), Manna *et al* (2005), and the present histopathological study showed pyknotic nuclei and vacuolated. Furthermore, the key components of the histopathologic change include congestion and bleeding. Cygon direct cytotoxic effects on endothelial cells might explain these findings (Alejandro *et al.*, 2012). Both severe congestion and hemorrhaging by damaging vascular walls and resulting in dilatation and congestion, oxidative stress and lipid peroxidation can cause blood artery congestion (Shady and Noor El-Deen, 2010). Cygon-induced oxidative stress may also result in degeneration.

ROS in various organs causes tissue damage (tissue atrophy, necrosis, exfoliation, and inflammation). This outcome is consistent with past toxicological investigations (Jeon *et al.*, 2013; Eman and Ghusoon, 2019). In vitro and in vivo, it might also help prevent or reduce oxidative damage to tissues and cells (Velmurugan *et al.*, 2002, Reifen *et al.*, 2004). Furthermore, degenerative alterations in the serosa, mucosa, and submucosal layers were seen, as well as localized necrosis, proliferation, and desquamation of the superficial villi (Eddleston and Clark 2011). Endosulfan exposure, causes alterations in the epithelial lining of the colon, indicating a disruption in intestinal absorption. Cengiz *et al.* (2001) discovered endosulfan-induced edema, degeneration, lymphocyte buildup in the lamina propria, pyknotic nuclei, and necrosis in the stomach of *Gambusia affinis*. After exposure to deltamethrin, the gut of fish showed infiltration of mononuclear leucocytes and eosinophils towards the lamina propria, as well as necrosis (Cengiz and Unlu, 2006). (Anwar, *et al.*, 2020) observed that organophosphorus insecticides such as Endosulfan increased pathogenicity in the gut and stomach due to the fact that organochloride generates organochloride acid in the presence of HCl released in the stomach, which has extremely corrosive qualities. This acid damages the mucous secreting cells of the gut lining, resulting in the abnormalities observed.

Endosulfan has also been related to pathological gastrointestinal symptoms such as mucosal lining damage, microvilli loss, cracked clay appearance of duodenal mucosa, and gastric mucosa with desquamated epithelial cells. As a result, it irritates and damages the intestinal mucous membrane, reducing absorption. Lead was thought to induce gastric ulcers by interfering with oxidative metabolism in the stomach, which increased the occurrence of gastric ulcers (Olaleye *et al.*, 2007).

The histopathological lesions in the intestine and stomach of animals receiving Aftovaxpur DOE vaccine were characterized by moderate lesions caused by the Aftovaxpur DOE vaccine innate immune response and provided partially protective immune responses against the Cygon effect on organs by a significant increase in IL-1 and IFN- $\gamma$  by DC (Petrovsky and Aguilar 2004). Other potential effectors include CD4<sup>+</sup> T-helper (Th) lymphocytes and cytotoxic CD8<sup>+</sup> T lymphocytes, which can stop the spread of infectious diseases by identifying and killing infected cells or by releasing particular antiviral cytokines. These Th cells may contribute to the body's defense by generating cytokines and supporting the development and upkeep of CD8<sup>+</sup> and B T-cell responses. Their primary cytokine production (interleukin [IL]-4 or interferon) led to the classification of effector CD4<sup>+</sup> T cells as either T-helper 1 (Th1) or T-helper 2 (Th2). This distinction was rendered obsolete upon the discovery that Th cells are classified into multiple subgroups, each possessing distinct cytokine-producing and homing characteristics. (Geginat *et al* 2014).

the possible long-term consequences of exposure to Cygon. causes irreversible histopathological alterations that result in long-term diseases like cancer or fibrosis; this finding is consistent with earlier research that demonstrated exposure to varying cygon dosages can cause cancer and liver fibrosis (Li *et al.*, 2021b). Notably, our earlier study showed that mice can develop colitis and colonic damage after 8 weeks of exposure to low-dose DLM (0.2 mg/kg/day) through the promotion of oxidative stress and disruption of intestinal flora (Ma *et al.*, 2023).

### Conclusions and Recommendation:

Based on the experimental findings of this study, we demonstrated that Cygon has a harmful toxic effect on mice, manifested in changes to the digestive system's histology and immunology. Based on the current study, we recommend that Cygon use be limited to a particular program.

### ACKNOWLEDGEMENTS

The authors are thankful to the head College of Veterinary Medicine, University of Baghdad for their assistance during work.

**NOVELTY STATEMENT**

The novelty of the study is focus on the immunotoxicity of Cygon as well as the histological status of stomach and intestine in mice.

**AUTHOR'S CONTRIBUTION**

Sabrin Ibraheem Mohsin, Salema Lafta Hassan, Taghred Jabbar Humadai, Designed and Performed the experiments, analyzed the data, contributed reagents, materials, analysis tools and wrote the paper.

**Conflict of interest**

The authors have declared no conflict of interest.

**REFERENCES**

- Alejandro Romero, Eva Ramos, Víctor Castellano, María Aranzazu Martínez, Irma Ares, Marta Martínez, María Rosa Martínez-Larrañaga, Arturo Anadón (2012). Cytotoxicity induced by deltamethrin and its metabolites in SH-SY5Y cells can be differentially prevented by selected antioxidants. *Sep*; 26(6): 823-30.
- Anwar H., Irfan S., Hussain G., Naeem Faisal M., Muzaffar H., Mustafa I., Mukhtar I., Malik S., Irfan Ullah M. Gut Microbiome: A New Organ System in Body. *Parasitol. Microbiol. Res.*, 2020; 1: 17–21.
- Baiyun RQ, Li SY, Liu BY, Lu J J, Lv YY, Xu, JW, Wu J.H, Li JY, Lv ZJ, Zhang ZG. Luteolin-mediated PI3K/AKT/Nrf2 signaling pathway ameliorates inorganic mercury-induced cardiac injury. *Ecotoxicol. Environ. Saf.*, 2018; 161: 655–661.
- Braunbeck, Appellbaum, T, Braunbeck S, Appellbaum. Ultrastructure alternation in the Liver and intestine of Crap Cyprinus carpio induced orally by ultra-low doses of Enodsulfan, *Dis. Aquat. Organ.*, 1999; 36: 183-200.
- Cengiz EI, Unlu E EI, Cengiz E. Sub lethal effects of commercial Deltamethrin on the structure of the gill, liver and gut tissues of Mosquito fish *Gambusia affinis*: a microscopic study, *Environ. Toxicol. Pharmacol.*, 2006; 21: 246-253. <http://doi.org/10.1016/j.etap.2005.08.005>
- Central Disease Control (2017.) CDC Zika Interim Response Plan; Central Disease Control: Atlanta, GA, USA.
- Chen Y, Huang TY, Shi W, Fang JS, Deng HK, Cui GZ. Potential targets for intervention against doxorubicin-induced cardiotoxicity based on genetic studies: a systematic review of the literature. *J. Mol. Cell. Cardiol.*, 2020; 138: 88–98.
- Eddleston M, Clark RF Insecticides – Organic phosphorus compounds and Carbamates. In: Goldfrank's toxicologic emergencies. 9th ed. New York: McGraw Hill, 2011; 1450–1457.
- El Okda E, Abdel-Hamid MA, Hamdy AM. Immunological and genotoxic effects of occupational exposure to cypermethrin pesticide. *Int. J. Occup. Med. Environ. Health.*, 2017; 30: 603–615. <http://doi.org/10.13075/ijomeh.1896.00810>
- Eman Hashim Yousif, Ghusoon Abdul Kareem Neamah Determination of Depleted Uranium Concentration and Histopathological Changes in Local Iraqi Fish and Chickens. *The Iraqi Journal of Veterinary Medicine*, 2019; 43(2): 90-101. <http://https://doi.org/10.30539/iraqijvm.v43i2.537>
- Gabbianelli R, Falcioni ML, Nasuti C, Cantalamessa F, Imada I, Inoue M. Effect of permethrin insecticide on rat polymorph nuclear neutrophils. *Chem. Biol. Interact.*, 2009; 182: 245–252. <http://DOI: 10.1016/j.cbi.2009.09.006>
- Geginat J, Paroni M, Maglie S, et al. Plasticity of human CD4 T cell subsets. *Front Immunol.*, 2014; 5: 630. <http://DOI: 10.3389/fimmu.2014.00630>
- House C, House JA Evaluation of techniques to demonstrate foot-and-mouth disease virus in bovine tongue epithelium: comparison of the sensitivity of cattle, mice, primary cell cultures, cryopreserved cell cultures and established cell lines. *Veterin Microbiol.*, 1989; 20(2): 99–109. [http://DOI: 10.1016/0378-1135\(89\)90033-3](http://DOI: 10.1016/0378-1135(89)90033-3)
- Huda S. Jassim Efficacy of Immune System Challenges with Tiny Enemy COVID-19. *The Iraqi Journal of Veterinary Medicine*, 2020; 44(1): 75-7975. <https://doi.org/10.30539/ijvm.v44i1.940>
- Hulse EJ, Haslam JD, Emmett SR, Woolley T. Organophosphorus nerve agent poisoning: managing the poisoned patient. *Br J Anaesth*, 2019; 123: 457–63.
- Ismail MF, Mohamed HM. Deltamethrin-induced genotoxicity and testicular injury in rats: comparison with biopesticide. *Food Chem. Toxicol.*, 2012; 50: 3421–3425. <https://doi.org/10.1016/j.fct.2012.07.060>
- Kamath V, Joshi AKR, Rajini PS. Dimethoate induced biochemical perturbations in rat pancreas and its attenuation by cashew nut skin extract. *Pestic Biochem Physiol.*, 2008; 90: 58–65 <https://doi.org/10.1016/j.pestbp.2007.07.007>
- Kumari A S, Kumar N S R Effects of water pollution on histology of intestine of two fresh water fishes from Hussainsagar lake (A. P.), *Indian J. Environ. Toxicol.*, 1997; 7: 68-70.
- Li S et al., 2021b. Exploring the liver fibrosis induced by deltamethrin exposure in quails and elucidating the protective mechanism of resveratrol. *Ecotoxicol. Environ. Saf.*, 2021; 211: 11501.
- Luma W Khalil, Layla Hashim Alol, Anwar I Obeid Effect of crude polyphenol extracted from black olive fruit (*Olea europaea*) on some physiological and immunological parameters in Males Rats Treated with Hydrogen Peroxide. *The Iraqi Journal of Veterinary Medicine*, 2013; 37(1): 83 – 89. <https://doi.org/10.30539/iraqijvm.v37i1.337>
- Luna LG. Manual of histologic staining methods of the armed forces institute of pathology, 3rd edn. McGraw-Hill, New York, NY, 1968; 55-78.
- Ma R, et al. Chronic exposure to low-dose deltamethrin can lead to colon tissue injury through PRDX1 inactivation-induced mitochondrial



- oxidative stress injury and gut microbial dysbiosis. *Ecotoxicol. Environ. Saf.*, 2023; 264: 115475.
23. MacDonald-Wicks LK, Wood LG, Garg M L. Review Methodology for the determination of biological antioxidant capacity in vitro: a review. *J. Sci. Food Agric.*, 2006; 86(13): 2046-2056. <http://DOI: 10.1002/jsfa>
  24. Maecker H. Cytotoxic T cell responses to DNA vaccination: dependence on antigen presentation via class II MHC. *Journal of Immunology*, 1998; 161: 6532-6536.
  25. Mahajan NK, Kulshreshtha RC, Malik G and Dahiya JP. Immunogenicity of major cell surface protein(s) of *Brucellamelitensis* Rev 1. *Journal Research of Veterinary research communications.*, 2005; 29(3): 189-199.
  26. Manna S, Bhattacharyya D, Mandal TK, Das S. Repeated dose toxicity of deltamethrin in rats. *Indian J Pharmacol.*, 2005; 37(3): 160-164. <http://DOI: 10.4103/0253-7613.16212>
  27. Olaley SB, Adaramoye OA, Erighali PP, Adeniyi OS. Lead exposure increases oxidative stress in the gastric mucosa of HCl/ethanol-exposed rats. *World J. Gastroenterol.*, 2007; 13: 5121-5216. <http://DOI: 10.3748/wjg.v13.i38.5121>
  28. Persis VT and Kalaiarasi JMV. Histopathological Responses of *Mystus vittatus* to Chronic Sublethal and Acute Lethal Toxicity of an Organophosphate Pesticide. *J. E xpt Zoo India.*, 2001; 4(1): 103- 108.
  29. Petrovsky N, Aguilar JC. Vaccine adjuvants: Current state and future trends. *Immunol Cell Biol.*, 2004; 82(5): 488-496. <http://DOI: 10.1111/j.0818-9641.2004.01272.x>
  30. Ramond A, Godin D, Ribuot C, Totson P. Oxidative stress Mediates cardiac infarction aggravation induced by intermittent hypoxia. *Fundam. Clin. Pharmacol.*, 2011; 1(10): 111-115. <http://DOI: 10.1111/j.1472-8206.2011.01015.x>
  31. Reifen R, Nissenkorn A, Matas Z, Bujanover Y. 5-ASA and lycopene decrease the oxidative stress and inflammation induced by iron in rats with colitis. *J. Gastroenterol.*, 2004; 39: 514-519. <http://DOI: 10.1007/s00535-003-1336-z>
  32. Sabina Tos-Luty, Agnieszka Haratym-Maj, Jadwiga Latuszynska, Daniela Obuchowska-Przebirowska, Malgorzata Tokarska-Rodak. Oral toxicity of deltamethrin and fenvalerate in Swiss mice. *Ann Agric Environ Med*, 2001; 8: 245-254.
  33. Saleh HM (1999). Immunological evaluation of the locally produced Brucellins in the sheep infected with *Brucella* and immunized with Rev 1 vaccine. Msc. Thesis. Vet. Med. Coll. Bagh. Univ.
  34. Shady A M, Noor El-Deen F I. Effect of Chlorpyrifos on Thyroid Gland on adult Male Albino Rats. *Egyptian Journal of Histology.*, 2010; 33(3): 441 – 450.
  35. SPSS (2008) Statistical Package for Social Science, SPSS Users Guide. Statistics Version 16. N.C. Usa.
  36. Su YY, Wei HD, Bi Y.J, Wang YN, Zhao P, Zhang R.X, Li X, Li JH, Bao J, Pre-cold acclimation improves the immune function of trachea and resistance to cold stress in broilers. *J. Cell. Physiol.*, 2019; 234: 7198-7212.
  37. Velmurugan B, Bhuvaneswari V, Nagini S. Antiperoxidative effects of lycopene during N-methyl-N-nitro-N-nitrosoguanidine-induced gastric carcinogenesis. *Fitoterapia.*, 2002; 73: 604-611. [http://DOI: 10.1016/s0367-326x\(02\)00216-2](http://DOI: 10.1016/s0367-326x(02)00216-2)
  38. Velmurugan B, Selvanayagam M, Cengiz EI, Unlu E. The effects of fenvalerate on different tissues of freshwater fish *Cirrhinus mrigala*. *J Environ Sci Health.*, 2007; 42: 157-163. <http://DOI: 10.1080/03601230601123292>
  39. Wei HD, Zhang RX, Su YY, Bi YJ, Li X, Zhang X, Li JH, Bao J. Effects of acute cold stress after long-term cold stimulation on antioxidant status, heat shock proteins, inflammation and immune cytokines in broiler heart. *Front. Physiol.*, 2018; 9: 1589.
  40. World Health Organization (WHO) (2015). Specifications and Evaluations for Public Health Pesticides. Permethrin (25:75 Cis:Trans Isomer Ratio, Nonracemic) 3-Phenoxybenzyl (1RS,3RS;1RS,3SR)-3-(2,2 dichlorovinyl)-2,2-dimethyl-cyclopropane Carboxylate; World Health Organization: Geneva, Switzerland.
  41. World Health Organization (WHO) (2017). Deltamethrin Long-Lasting (Coated onto Filaments) Insecticidal Net(s)-Cyano-3-phenoxybenzyl (1r,3r)-3-(2,2dibromovinyl)-2,2-dimethylcyclopropane Carboxylate. World Health Organization: Geneva, Switzerland. <https://extranet.who.int/pqweb/vector-control-products>