

# WORLD JOURNAL OF ADVANCE HEALTHCARE RESEARCH

SJIF Impact Factor: 6.711

ISSN: 2457-0400 Volume: 9 Issue: 3 Page N. 29-35 Year: 2025

Original Article

www.wjahr.com

# POSSIBLE ROUTES OF BACTERIAL TRANSMISSION; A STUDY ON TRICYCLES USED WITHIN ABAKALIKI METROPOLIS

Odo Ikechukwu Ituma,<sup>1</sup> Udu-Ibiam Onyinyechi E,<sup>2</sup> Anger Robin A,<sup>3</sup> Ovuaja Maureen N.<sup>4</sup>

<sup>1</sup>Alex ekwueme federal university Ndufu Alike Ikwo.
<sup>2</sup>Ebonyi State University, Abakaliki.
<sup>3</sup>University of Agriculture Markudi Benue state.
<sup>4</sup>David Umahi federal University of medical Science Uburu.

Article Received date: 23 December 2024Article Revised date: 13 January 2024Article Accepted date: 03 February 2025



\*Corresponding Author: Udu-Ibiam, Onyinyechi E.

Alex Ekwueme FEDERAL University Ndufu Alike Ikwo.

# ABSTRACT

Strategic places in tricycles used in Abakaliki metropolis were aseptically swabbed and examined using standard microbiological method. The bacterial count revealed that the examined tricycles analyzed had a bacterial load that ranges from  $3.2 \times 10^7$  to  $1.1.2 \times 10^7$ . Morphological and biochemical characteristics of bacteria isolates from the samples shows that a total of three isolates were identified and they includes; *Staphylococcus aureus, Eschericia coli* and *Klebsiella* spp. Predominantly *Staphylococcus aureus* was isolated from most of the tricycles examined at a percentage of 38(80%) whereas the least was 1(2%). When the isolates were tested against the local disinfectant it was observed that Germicide  $Z^{\odot}$  had a greater inhibitory effect on the isolates closely followed by De<sup>©</sup> showing an inhibition zone diameter that ranged between 12mm to 40mm and 20 to 40 mm respectively. Disinfectant IZ<sup>®</sup> had no inhibitory effect on the isolates.

KEYWORDS: Tricycles, Fomites, Transmission, Infectious Diseases, Disinfectant, Antiseptics.

#### **INTRODUCTION**

An infection is the invasion of tissues by pathogens, their multiplication, and the reaction of host tissues to the infectious agent and the toxins they produce (Acevedo et 2019). An infectious disease, also known as al.. a transmissible disease or communicable disease which is an illness resulting from an infection. Infections can be caused by a wide range of pathogens, most prominently bacteria and viruses (Adams et al., 2015). Hosts can fight infections using their immune systems. Mammalian hosts react to infections with an innate response, often involving inflammation, followed by an adaptive response. In certain cases, infectious diseases may be asymptomatic in a given host. In the latter case, the disease may only be defined as a "disease" (which by definition means an illness) in hosts secondarily become ill after contact with who an asymptomatic carrier. An infection is not synonymous with an infectious disease, as some infections do not cause illness in a host (Adam et al., 2013).

As bacterial and viral infections can both cause the same kinds of symptoms, it can be difficult to distinguish which is the cause of a specific infection (Abdulwasiu et

al., 2022). Distinguishing the two is important, since viral infections cannot be cured by antibiotics whereas bacterial infections can (AAMC, 2015).

There is a general chain of events that applies to infections, sometimes called the chain of infection (Brown, 1987) or transmission chain. The chain of events involves several steps - which include the infectious agent, reservoir, entering a susceptible host, exit and transmission to new hosts. Each of the links must be present in a chronological order for an infection to develop. Understanding these steps helps health care workers target the infection and prevent it from occurring in the first place (CDC, 2023). Infection begins when an organism successfully enters the body, grows and multiplies. This is referred to as colonization. Most humans are not easily infected. Those with compromised or weakened immune systems have an increased susceptibility to chronic or persistent infections. Individuals who have a suppressed immune system are particularly susceptible to opportunistic infections. Entrance to the host at host-pathogen interface, generally occurs through the mucosa in orifices like the oral cavity, nose, eyes, genitalia, anus, or the microbe can enter through open wounds. While a few organisms can grow at the initial site of entry, many migrate and cause systemic infection in different organs. Some pathogens grow within the host cells (intracellular) whereas others grow freely in bodily fluids. Wound colonization refers to non-replicating microorganisms within the wound, while in infected wounds, replicating organisms exist and tissue is injured (CDC, 2019). All multicellular organisms are colonized to some degree by extrinsic organisms, and the vast majority of these exist in either a mutualistic or commensal relationship with the host. the An example of former is the anaerobic bacteria species, which colonizes the mammalian colon, and an example of the latter are the various species of staphylococcus that exist on human skin. Neither of these colonizations is considered infections. The difference between an infection and colonization is often only a matter of circumstance. Non-pathogenic organisms can become pathogenic given specific conditions, and even the most virulent organism requires certain circumstances to cause a compromising infection. Some colonizing bacteria, such as Corynebacteria sp and Viridans streptococci, prevent the adhesion and colonization of pathogenic bacteria and thus have a symbiotic relationship with the host, preventing infection and speeding wound healing. The variables involved in the outcome of a host becoming inoculated by a pathogen and the ultimate outcome include: the route at which the pathogen enter the host to initiate infection, the internal virulence of the particular organism, the microbial load of the organism implicated, the immune status of the host in question. As an example, several staphylococcal species remain relatively non infective on the skin, but, when present in a normally sterile region, such as in the capsule of a joint or the peritoneum, it multiplies rapidly without resistance and cause harm (CDC, 2019).

Disease can arise if the host's protection and immune statues mechanisms are compromised and the organism inflicts damage on the host. Microorganisms can cause lots of damages on the tissues by releasing a variety of toxins or destructive enzymes. For example, Clostridium tetani releases a toxin that paralyzes muscles, and staphylococcus releases toxins that produce shock and sepsis. Not all infectious agents have the capacity to cause disease in all hosts. For example, less than 5% of individuals infected with polio develop disease (Ericson et al., 1999; Wayback machine, 2010). On the other hand, some infectious agents are known to be highly virulent. The prion causing mad cow disease and Creutzfeldt-Jakob disease is known to kills all animals and people that are infected(Duerkop and Hooper, 2013; Negut et al., 2018). Some of these virulent infections can be persistent, especially when the host is continuously being exposed to the infectious agents through fomites.

Persistent infections occur because the body is unable to expel the organism after the initial infection. Persistent

infections are characterized by the continual presence of the infectious organism, often as latent infection with occasional recurrent relapses of active infection. There are some viruses that can continually cause infection by infecting different cells of the body. Some viruses once acquired never leave the body. A typical example is the herpes virus, which tends to hide in nerves and become reactivated when specific circumstances arise (Hector and Booksmythe, 2019; Hotez et al., 2006).

Persistent infections cause millions of deaths around the world each year (CDC, 2019). Chronic infections by parasites shares for a high morbidity and mortality in many underdeveloped countries (CDC, 2019).

For infecting organisms to survive and repeat the infection cycle in other hosts, they (or their progeny) must leave an existing reservoir and cause infection elsewhere. Infection transmission can take place via many known likely routes (GBC, 2014; Microbiology book. Org, 2017) either by; Droplet contact, also known as the respiratory route, (Clevland Clinic 2022; NIPA, 2023) and the resultant infection that results is called airborne disease. Tricycles are the most common means of transporting persons and goods within abakaliki, in Ebonyi state. People from all works of life patronize tricycles on daily basis. From the early hours to late ours of the day. This serves as a good means of transferring infectious agent from a carrier to an unsuspecting individual. Fecal-oral transmission, is another means of disease transmission wherein foodstuffs or water become contaminated either (by people not washing their hands before preparing food, or sewage not properly treated going into a drinking water supply) and the people who and drink them become infected. Sexual eat transmission, with the result being called sexually transmitted infection, Oral transmission, diseases that are transmitted primarily by oral means may be caught through direct or indirect contact such as by sharing a drinking glass or a cigarette (Mada and Alam, 2019; NIH, 2007).

# MATERIAL AND METHODS

# Instruments

The following equipment/instrument was used: Autoclave (Olympic company), Microscope (Olympic company), wire loop, Bunsen burner, autoclave, oven, incubator, weighing balance, spatula.

# **Chemicals and Reagents**

The following reagents and chemicals that were used in this study includes: sterile water, Dimethyl Sulphur Oxide, normal saline.

# Media

The following media were used for this research work, Nutrient broth, Nutrient agar and Muller Hinton aga Eosin methylene Blue agar, mannitol salt, Macconkey agar, salmonella-shigella agar. All media were prepared aseptically according to the manufacturers' instruction.

#### Disinfectants

The following disinfectants were used in this work,  $De^{\circ}$ ,  $IZ^{\circ}, Z^{\circ}$ .

#### Study Area

The study area is Abakaliki town in Ebonyi State located in south eastern part of Nigeria. It is located 64 kilometers southeast of Enugu. Abakaliki is situated on latitude  $6^{0}20$ 'N and longitude  $8^{0}06E$ .

#### Sample Collection

Swabs were taken from sensitive area in the commercial tricycles used in abakaliki metroplolis (handles). The samples were transported to the microbiology laboratory of EBSU, Abakaliki where they were analyzed following standard techniques in Microbiology (Anie et al., 2017).

#### 3.3 Analysis of Samples

Samples were streaked on freshly prepared sterile media after undergoing a tenfold serial dilutions. They were all incubated for 18 to 24 hours at  $37^{0}$ c (AOAC, 1995).

#### **Determination of Aerobic Plate Count**

Standard plate count method proposed by AOAC, (2004) was used to determine the total aerobic colony count of the samples. Only plates with moderate growth were

Table 1: Colony count from commercial Tricycles.

counted. The average microbial loads of the samples obtained were expressed as colony forming units per Milliliter (Cfu/ml).

# Isolation and Identification of Bacteria Isolated from Tricycles

Mac Conkey agar, Eosin methylene Blue agar, mannitol salt and salmonella – shigella agar were employed for the isolation of bacteria for the purpose of identification. Mac Conkey agar was used to isolate lactose fermenting gram negative bacteria, Eosin methylene Blue agar was used for the selective isolation of enteric coliforms, mannitol salt agar was used for the selective isolation of salt-tolerant bacteria and salmonella – shigella agar was used for the isolation of enteric bacilli particularly *Salmonella and Shigella* species. All plates were incubated at  $37^{\circ}$ C for 24 hours. Identification of bacteria isolates was based on the standard culture, morphological and biochemical methods (CLSI, 2015).

# RESULTS

Out of the fifty (50) tricycles sampled only thirty Seven (37) had visible growth. The colony count is as shown in Table 1 below. The bacterial count revealed that the examined tricycles analyzed had a bacterial load that ranges from  $3.2 \times 10^7$  to  $1.1.2 \times 10^7$ .

S/no	Sample code	Colony count/colony forming unit
1	K1	16 (3.2x 10 <sup>4</sup> )
2	K4	48 (9.6 x 10 <sup>4)</sup>
3	K6	$56 (1.12 \times 10^5)$
4	K7	$104(2.08 \text{ x } 10^5)$
5	K8	80(1.6 x 10 <sup>5</sup> )
6	K9	$56(1.12 \text{ x}10^5)$
7	K10	$68 (1.36 \times 10^5)$
8	K11	$112 (2.24 \text{ x} 10^5)$
9	K12	$88 (1.76 \times 10^5)$
10	K13	92 ( $1.84 \times 10^5$ )
11	K14	96 ( $1.92 \times 10^5$ )
12	K15	$54 (1.08 \text{ x} 10^5)$
13	K16	$119(2.38 \times 10^5)$
14	K17	$86 (1.72 \times 10^5)$
15	K18	$45(9.0 \times 10^4)$
16	K19	$43(8.6 \times 10^4)$
17	K20	$23(4.6 \times 10^4)$
18	K21	$54 (1.08 \times 10^5)$
19	K23	$64 (1.28 \text{ x} 10^5)$
20	K27	98 ( $1.96 \times 10^5$ )
21	K28	$105 (2.10 \times 10^5)$
22	K30	$26 (5.2 \times 10^4)$
23	K31	$28 (5.6 \times 10^4)$
24	K32	$80 (1.6 \times 10^5)$
25	K33	$56 (1,12 \text{ x}10^5)$
26	K34	$68 (1.36 \times 10^5)$
27	K35	$112 (2.24 \text{ x}10^5)$
28	K36	$88 (1.76 \times 10^5)$
29	K37	92 $(1.84 \times 10^5)$
30	K38	96 $(1.92 \times 10^5)$

I

31	K40	$115(2.3 \times 10^5)$
32	K41	98 (1.96 x10 <sup>5</sup> )
33	K45	$45 (9.0 \times 10^5)$
34	K47	43 (8.6 x 10 <sup>5</sup> )
35	K48	$23 (4.6 \times 10^4)$
36	K49	$54 (1.08 \times 10^5)$
37	K50	$64(1.28 \times 10^5)$

The percentage distribution of isolates from the commercial tricycles was determined as shown in Table 2 below. *S. aureus* was the highest with 35(77.7)

followed by *Eschericia coli* with value 9(20). The least was *Klebsiella pneumonia* with 1(2.22).

Table 2: Percentage distribution of isolates from the Commercial Tricycles.

S/No	ISOLATES FROM MOBILE PHONES	PERCENTAGE DISTRIBUTION
1	S. aureus	35(77.7)
2	Klebsiella pneumonia	1(2.22)
3	Eschericia coli	9(20)
	TOTAL: 45	

The inhibition Inhibition Zone Diameter of Isolates from Commercial Tricycle with  $IZ^{\odot}$  was determined. The

entire isolates were resistant to this particular dis infectant, this result is as shown in Table 3 below.

Tuble 5, Innibition Lone Dianteer of 1901ates from Commercial freque (film 12)	Table 3: Inhibition	a Zone Diameter	<b>OF</b> Isolates from	Commercial Tricy	ycle With IZ <sup>©</sup> .
--------------------------------------------------------------------------------	---------------------	-----------------	-------------------------	------------------	-----------------------------

Isolates	Inhibition zone concentration	25	
S. aureus	Nil	Nil	Nil
E. coli	Nil	Nil	Nil
K. pneumonia	Nil	Nil	Nil

In Table 4 below, it shows the result of the isolates against another disinfectant  $DE^{\odot}$ . It was observed that the antibacterial agent was able to inhibit the growth of the isolates giving the highest inhibition zone diameter of 40mm at 100mglml and 25mm at 50mg/ml for *S. aureus*.

For E. coli 100mg/ml concentration gave an IZD of 20mm whereas 25mg/ml equally showed an IZD of 20mm. *K. pneumoniae* had the same value for 100mg/ml and 50mg/ml which is 20mm whereas 25mg/ml exhibited an IZD of 25mm.

Table 4: Inhibition Zone Diameter Of Isolates from Commercial Tricycle With De	e <sup>©</sup> .
--------------------------------------------------------------------------------	------------------

Isolates	Inhibition zone concentration (	diameter (mm)/ (mg/ml) 100 50	25
S. aureus	40	25	30
E. coli	20	23	20
K. pneumonia	20	20	25

In Table 5 below, it shows the result of the isolates against another disinfectant  $Z^{\odot}$ . It was observed that the antibacterial agent was able to inhibit the growth of the isolates giving the highest inhibition zone diameter of 40mm at 100mglml and 25mm at 50mg/ml for *S. aureus*.

For E. coli 100mg/ml concentration gave an IZD of 20mm whereas 25mg/ml equally showed an IZD of 35mm. *K. pneumoniae* had the value of 33mm for 100mg/ml and 50mg/ml the IZD value was 15mm whereas 25mg/ml exhibited an IZD of 12mm.

Table 5: Inhibition Zone Diameter OF Isolates from Commercial Tricycle Z  $^{\circ}$ .

Isolates	Inhibition zone di concentration (m	25	
S. aureus	40	25	19
E. coli	20	35	25
K. pneumonia	33	15	12

#### DISCUSSION

This research work examined 50 tricycles that ply the Abakaliki metropolis to determine possibility of transmission of diseases through fomites like tricycle.

I

Out of the 50 tricycle examined only 37 tricycles had visible growth. The total viable count ranged from  $3.2 \times 10^7$  to  $1.1.2 \times 10^7$ . Morphological and biochemical characteristics of bacteria isolates from the samples after

comparison with the Bergey's Determinative Bacteriology Manual (1994) shows that a total of three isolates were identified and they include; *Staphylococcus* Eschericia coli and Klebsiella aureus spp. Predominantly Staphylococcus aureus was isolated from most of the tricycles examined. This is observed from the result of the percentage distribution of isolates from the commercial tricycles, S. aureus was the highest with 35(77.7) followed by Eschericia coli with value 9(20). The least was *Klebsiella pneumonia* with 1(2.22). This result obtained is similar to that carried out by Abdulwasiu et al (2022) on handles of doors from buildings. Similar research reported found out that 84.6% of isolates from faucets examined were gram positive cocci with only 15.4%-gram negative cocci. Vehicle transmission by an inanimate reservoir (food, water, soil)(Acevedo et al., 2019) has been equally reported by some credible researchers which mainly washing one's hands appears to be an effective way to prevent the spread of infectious disease(Reddy et al., 2012), wearing gowns, and wearing face masks can help prevent infections from being passed from one person to another. Aseptic technique was introduced in medicine and surgery in the late 19th century and greatly reduced the incidence of infections caused by surgery. Frequent hand washing remains the most important defense against the spread of unwanted organisms. The isolates from the tricyles were tested against locally known and used disinfectant to determine its efficacy against the bacterial isolates. The result of the isolates against disinfectant DE<sup>©</sup> showed that the antibacterial agent was able to inhibit the growth of the isolates giving the highest inhibition zone diameter(IZD) of 40mm at 100mglml and 25mm at 50mg/ml for S. aureus. For E. coli 100mg/ml concentration gave an IZD of 20mm whereas 25mg/ml equally showed an IZD of 20mm. K. pneumoniae had the same value for 100mg/ml and 50mg/ml which is 20mm whereas 25mg/ml exhibited an IZD of 25mm. The result of the isolates against another disinfectant Z<sup>©</sup>. It was observed that the antibacterial agent was able to inhibit the growth of the isolates giving the highest inhibition zone diameter of 40mm at 100mglml and 25mm at 50mg/ml for S. aureus. For E. coli 100mg/ml concentration gave an IZD of 20mm whereas 25mg/ml equally showed an IZD of 35mm. K. pneumoniae had the value of 33mm for 100mg/ml and 50mg/ml the IZD value was 15mm whereas 25mg/ml exhibited an IZD of 12mm. This finding is in line with Olowe et al., (2004) that observed inhibitory effect on some of the disinfectants examined. Disinfectant IZ<sup>©</sup> had no inhibitory effects on the isolates from the tricycles. This is likely to be a case of bacterial resistance to the disinfectant used. Antimicrobial substances in form of antiseptics, (which are applied to living tissue/skin), disinfectants( which destroy microorganisms found on non-living objects), antibiotics, called prophylactic when given as prevention rather as treatment of infection. However, long term use of antibiotics leads to resistance of bacteria. While humans do not become immune to antibiotics, the bacterium does become immune to

antibiotics as it develops resistance to the antibacterial agent. Thus, avoiding the usage of antibiotics longer than necessary helps to prevent bacteria from forming mutations that aide in antibiotic resistance (Peterson, 1990; Pinsky and Hayden, 2019).

This findings has further showed that indeed fomites harbor bacteria and can serve as a means of transferring infectious materials but constant cleaning of the surfaces of this commercial tricycle can prevent some communicable diseases and hence control epidemic in the long run.

# REFERENCES

- Abdulwasiu O.H., Obeagu, E & Onu, F.U. A Survey of microbial contamination of door handles in various locations in Lokoja metropolis, Kogi state, Nigeria. Int. J. Curr. Res. Biol. Med., 2022; 7(1): 8-16.
- Acevedo MA, Dillemuth FP, Flick AJ, Faldyn MJ, Elderd BD. "Virulence-driven trade-offs in disease transmission: A meta-analysis". Evolution, 2019; **73**(4): 636–647. doi:10.1111/evo.13692. PMID 30734920. S2CID 73418339. Archived from the original on 2022-12-04. Retrieved 2022-06-28.
- Adams, R. I., Bateman, A. C., Bik, H. M., & Meadow, J. F. Microbiota of the indoor environment: a meta-analysis. Microbiome, 2015; 3(1): 49.
- Adams, R. I., Bhangar, S., Pasut, W., Arens, E. A., Taylor, J. W., Lindow, S. E., Bruns, T. D. Chamber bioaerosol study: outdoor air and human occupants as sources of indoor airborne microbes. PloS one, 2015; 10(5).
- Adams, R. I., Miletto, M., Taylor, J. W., & Bruns, T. D. Dispersal in microbes: fungi in indoor air are dominated by outdoor air and show dispersal limitation at short distances. The ISME journal, 2013; 7(7): 1262-1273.
- Anie, C. O. . Ugwu, M. C., Ibezim E. C. and. Esimone, C. O Antibiogram of Methicillin-Resistant Staphylococcus aureus Isolates among Healthy Human Subjects in Oleh, South-Southern Nigeria. *International journal of Current Microbiological and Applied Science*, 2017; 6(9): 3710-3716.
- Association of Official Analytical Chemists (AOAC). Official analytical chemistry (1995). International Journal of Innovative Science and Research Technology, 20th ed, USA., 1995; 1058–1059.
- 8. Association of Official Analytical Chemists (AOAC), Official methods of analysis of the association, 2004; 7(8): 31-33.
- Association of American Medical Colleges. (2015) "Infectious Disease, Internal Medicine". Archived from the original on 2015-02-06. Retrieved 2015-08-20. Infectious disease is the subspecialty of internal medicine dealing with the

diagnosis and treatment of communicable diseases of all types, in all organs, and in all ages of patients.

- 10. Bergey's Determinative Bacteriology Manual, 1994.
- Brown PJ. "Microparasites and Macroparasites". Cultural Anthropology, 1987; 2 (1): 155–71. doi:10.1525/can.1987.2.1.02a00120. JSTOR 656401
- 12. Centers for Disease Control and Prevention (2023) "How Infections Spread". Archived from the original on 2 June 2023. Retrieved 17 October 2021.
- 13. Centers for Disease Control and Prevention (2019). "Types of Fungal Diseases". . Archived from the original on 2020-04-01. Retrieved 2019-12-09.
- 14. Centers for Disease Control and Prevention. (2019). "About Parasites". Archived from the original on 2019-12-25. Retrieved 2019-12-09.
- 15. Cleveland Clinic (2022). "Runny Nose: Symptoms, Causes & Treatment". Archived from the original on 2022-05-10. Retrieved 2022-04-22.
- 16. Clinical and Laboratory Standard Institute (CLSI), Performance standards for antimicrobial disk susceptibility tests; Approved standard- 12th ed CLSI document M02-A12. Wayne, PA: Clinical and Laboratory Standards Institute, 2015; 31-37.
- Duerkop BA, Hooper LV. "Resident viruses and their interactions with the immune system". Nature Immunology, 2013; 14(7): 654–59. doi:10.1038/ni.2614. PMC 3760236. PMID 2377879 2.
- Ericson L, Burdon JJ, Müller WJ. "Spatial and temporal dynamics of epidemics of the rust fungus Uromyces valerianae on populations of its host Valeriana salina". Journal of Ecology, 1999; 87(4): 649–658.
  Bibcode:1999JEcol..87..649E. doi:10.1046/j.1365-

2745.1999.00384.x. S2CID 86478171.

- 19. GBD (2014). Mortality and Causes of Death Collaborators . "Global, regional, and national agesex specific all-cause and cause-specific mortality for 240 causes of death, 1990–2013: a systematic analysis for the Global Burden of Disease Study".
- Hector TE, Booksmythe I. "Digest: Little evidence exists for a virulence-transmission trade-off". Evolution, 2019; 73(4): 858–859. doi:10.1111/evo.13724. PMID 30900249. S2CID 85 448255.
- 21. Hotez PJ, Bundy DA, Beegle K, Brooker S, Drake L, de Silva N, Montresor A, Engels D, Jukes M (2006), Jamison DT, Breman JG, Measham AR, Alleyne G (eds.), "Helminth Infections: Soil-Helminth Infections transmitted and Schistosomiasis", Disease Control Priorities in Developing Countries (2nd ed.), Washington (DC): Bank, ISBN 978-0-8213-6179-5, World PMID 21250326, archived from the original on 2016-10-10, retrieved 2021-08-13.
- 22. Infection Cycle "Infection Cycle Symptoms and Treatment". Archived from the original on 2023-11-10. Retrieved 2023-11-10.
- 23. Mada PK, Jamil RT, Alam MU (2019), "Cryptococcus (Cryptococcosis)",

StatPearls, StatPearls Publishing, PMID 28613714, archived from the original on 2020-06-19 retrieved 2019-12-09.

- 24. Microbiologybook.org "Clinical Infectious Disease Introduction". Archived from the original on 2017-04-20. Retrieved 2017-04-19.
- 25. Mideo N, Alizon S, Day T. "Linking within- and between-host dynamics in the evolutionary epidemiology of infectious diseases". Trends in Ecology & Evolution, 2008; 23(9): 511–517. doi:10.1016/j.tree.2008.05.009. PMID 18657880.
- 26. Mordecai EA, Cohen JM, Evans MV, Gudapati P, Johnson LR, Lippi CA, Miazgowicz K, Murdock CC, Rohr JR, Ryan SJ, Savage V, Shocket MS, Α. Thomas Stewart Ibarra MB. Weikel DP. "Detecting the impact of temperature on transmission of Zika, dengue, and chikungunya using mechanistic models". PLOS Neglected Tropical Diseases, 2017; 11(4): e0005568. doi:10.1371/journal.pntd.0005568. PMC 5423694. P MID 28448507.
- 27. National Institutes of Health (US), Study BS (2007), "Understanding Emerging and Re-emerging Infectious Diseases", NIH Curriculum Supplement Series [Internet], National Institutes of Health (US), archived from the original on 2023-06-26, retrieved 2023-11-17.
- Negut I, Grumezescu V, Grumezescu AM. "Treatment Strategies for Infected Wounds". Molecules, 2018; 23(9): 2392. doi:10.3390/molecules23092392. ISSN 1420-3049. PMC 6225154. PMID 30231567.
- 29. NIPA (2023). Bacteria Bacterial vs. Viral infections". www.antibioticsinfo.org. Archived from the original on 2023-11-10. Retrieved 2023-11-10.
- Olowe, O.A., Olayemi A.B., Eniola KIT., Adeyeba O.A. Antibacterial activity of some selected disinfectants regularly used in hospitals, *African journal of clinical and experimental microbiology*, 2004; 5(1): 126-130.
- 31. Onaolapo J.A. Contamination of hospitals disinfectants and antiseptics. *Pharmacy world journal*, 1990; 74: 118-119.
- Peterson JW (1996). Bacterial Pathogenesis. University of Texas Medical Branch at Galveston. ISBN 9780963117212. PMID 21413346. Archived f rom the original on 2016-04-25. Retrieved 2022-10-20.
- Pinsky BA, Hayden RT. "Cost-Effective Respiratory Virus Testing". Journal of Clinical Microbiology, 2019; 57(9): e00373–19. doi:10.1128/JCM.00373-19. ISSN 0095-1137. PMC 6711893. PMID 31142607.
- 34. Reddy M, Gill SS, Wu W. "Does this patient have an infection of a chronicwound?". *JAMA*, 2012; 307(6): 605–11. doi:10.1001/jama.2012.98. PMID 22318282.
- 35. Wayback Machine. Intestinal Parasites and Infection Archived 2010-10-28 fungusfocus.com –

Retrieved on 2010-01-21 *Lancet*, 2010; **385**(9963): 117–71. doi:10.1016/S0140-6736(14)61682-2. PMC 4340604. PMID 25530442.