

ESTABLISHED LEAF FLAVONOIDS AS PHYTOLOGANDS FROM *ANDROGRAPHIS PANICULATA* (BURM. F.) WALL. EX NEESFOR ANTIBACTERIAL ACTIVITY AGAINST BACTERIAL DNA-GYRASE B RECEPTOR: AN *IN SILICO* APPROACH

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Received date: 26 June 2020

Revised date: 16 July 2020

Accepted date: 06 August 2020

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ABSTRACT

The medicinal plant, *Andrographis paniculata* (Burm. f.) Wall. Ex Nees is a common herbal plant having several medicinal properties in the leaves of plant. The medicinal properties are depending upon by the presence of several phytochemicals. Among different medicinal values, potent antibacterial properties are known as natural antibacterial agents. The objective of the present study was to detect receptor-ligand binding energy and interaction through molecular docking for lead phytochemicals established in the leaf of *A. paniculata* against bacterial DNA gyrase B protein (PDB ID: 3G7B). Molecular docking was performed by using software (iGEMDOC, Version 2.1) to detect binding energy and interaction. The interaction with residues was visualized in the AutoDoc tool (Version 1.5.6). Among 9 phytochemicals and 1 antibiotic (Ciprofloxacin), highest binding energy value was obtained in 5-hydroxy-7,8,2',5'-tetramethoxy-flavone (-121.862 Kcal/mol) when compared to Ciprofloxacin (-106.621 Kcal/mol). The binding interaction of target protein with this phytochemical found binding at the active site due to competitive inhibition. In conclusion, phytochemical 5-hydroxy-7,8,2',5'-tetramethoxy-flavone can be alternative of synthetic antibacterial drug as per binding energy value and residue interaction. It is suggesting further pharmacological and toxicological assay with this phytochemical after isolation from medicinal plant (*A. paniculata*).

**KEYWORDS:** Medicinal plant; *A. paniculata*; Phytochemicals; Bacterial DNA gyrase B; Molecular docking; Receptor-ligand binding; *In silico* prediction.

INTRODUCTION

The medicinal herb, *Andrographis paniculata* (Burm. f.) Wall. Ex Nees is a common plant species containing several phytochemicals in the leaves of plant to prevent different diseases. Among several diseases, natural product from this plant is showing research interest to detect lead compounds for the prevention of bacterial infection. On the other hand, it was observed that bacterial resistance occurred by synthetic antibiotic during the prevention of infection but phytochemicals of plant species can be suitable antibacterial drugs and prevent infection without bacterial resistance<sup>[1,2,3,4]</sup>

It has been studied by many researchers that different form of crude plant extract of *A. paniculata* prevent bacterial infection caused by *Salmonella* sp.,

*Shigella sonnei*, *Escherichia coli*, gram A streptococci, and *Staphylococcus aureus*, *Pseudomonas aeruginosa*, methicillin resistant *S. aureus*, *Salmonella typhimurium*, *Streptococcus pneumoniae*, *Streptococcus pyogenes*, *Legionella pneumophila*, *Bordetella pertussis*, etc.<sup>[5,6,7]</sup> In present scenario it is interesting research to predict the lead compound(s) as single or combination of phytochemical(s) through virtual screening. Furthermore, experimentation with several phytochemicals after extraction from leaves is a tedious job along with time and cost involvement, etc.

In this regard, computer-based receptor-ligand binding as a suitable approach for structure-based drug or biopesticide screening and exact phytochemical or combinations of few phytochemicals can be predicted within a few hours by using computational

simulation.<sup>[8]</sup> It is well-known that the molecular docking tool is used to predict the interaction between a small molecule (ligand) and a macromolecule (protein) that describes the behavioural characterization of small molecules in the binding site of target receptor. From past to recent research, several researchers have been reported the importance of molecular docking.<sup>[8-21]</sup>

In recent research natural products are potential for the development of biopesticides. Antibiotic is an important drug to prevent various bacterial infection in plants and animals. Several researchers are investigating the inhibitory activities to prevent the multiplication of pathogenic bacteria. Among several enzymes in bacteria, DNA gyrase enzyme is most effective for metabolic regulation in bacteria and help in the process of DNA replication, and called as type II topoisomerase, which enhance bacterial multiplication.<sup>[22-25]</sup>

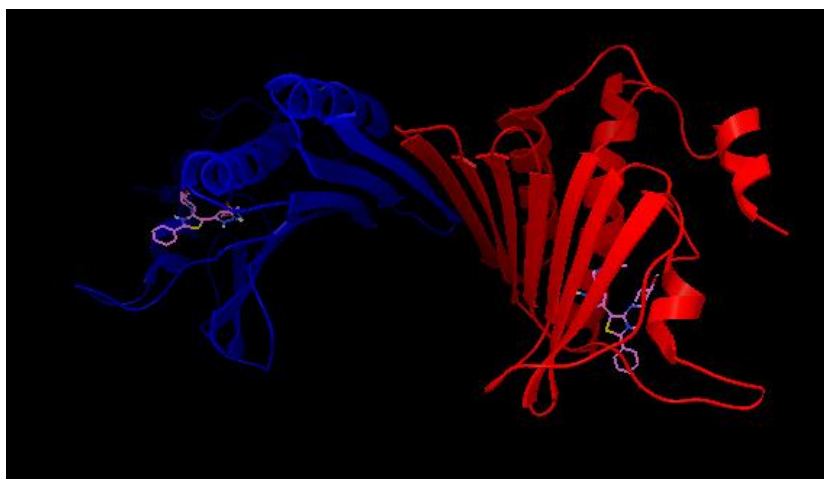
The present objective was to detect suitable receptor-ligand binding energy and molecular interaction through

molecular docking approach for phytochemicals established in *A. paniculata* against bacterial DNA gyrase B protein (PDB ID: 3G7B).

## MATERIALS AND METHODS

### Protein or receptor selection

The crystal three-dimensional (3-D) structure of receptor known as bacterial DNA gyrase B (PDB ID: 3G7B) was retrieved from protein data bank (<http://www.rcsb.org/>) as per validation report. After experimentation,<sup>26</sup> have deposited the X-ray diffraction crystallographic structure of the bacterial DNA gyrase B (Staphylococcus aureus gyrase B co-complex with methyl ({5-[4-(4-hydroxypiperidin-1-yl)-2-phenyl-1,3-thiazol-5-yl]-1H-pyrazol-3-yl}methyl) carbamate inhibitor) at 2.30 Å resolution. The 3-D ribbon structure was visualized in AutoDock tool developed by The Scripps Research Institute (Morris *et al.*, 1998)<sup>27</sup> and is exhibited in Fig 1.



**Fig. 1:** 3-D ribbon structure of DNA gyrase B (PDB ID: 3G7B) [Chain A = blue colour attached with ligand (ball and stick structure in green) at 471 position and Chain B = red colour attached with ligand (ball and stick structure in green) at 472].

### Phyto and synthetic ligands selection

The selection of phytochemicals (ligands) were done from the literatures as per antibacterial properties (Chen *et al.*, 2006; Jarukamjorn and Nemoto, 2008)<sup>28,29,30</sup>. In the present study, established 9 phytochemicals (flavonoids) of *A. paniculata* reported as antibacterial agents and one synthetic compound as known antibiotic (ciprofloxacin) were taken. The Canonical SMILES of these compounds were retrieved from the PubChem database ([www.ncbi.nlm.nih.gov/pubchem](http://www.ncbi.nlm.nih.gov/pubchem)) and .pdb file of each phytochemical was obtained from CORINA online server ([www.mn-am.com/](http://www.mn-am.com/)).

### Molecular docking and interaction study

The molecular docking was performed to know target lead by using iGEMDOCK (Version 2.1) developed by Yang and Chen (2004)<sup>31</sup>. The receptor-ligand interaction provides the values of binding energy, electrostatic, hydrogen-bonding and van der Waals after

completed docking. The above-mentioned profiles and compound structures were obtained by drug screening method for selected ten ligands as per genetic algorithm parameter by setting population size of 200 along with 70 generations and 3 solutions. After the completion of the docking, the post docking analysis was performed to find the docking pose and its energy values. The molecular docking was visualized by using AutoDoc tool, developed by The Scripps Research Institute and the results of three-dimensional structure were depicted. Docking of 9 phytochemicals and ciprofloxacin (ligand) with bacterial DNA gyrase B protein (PDB ID: 3G7B) were analysed to detect suitable binding energy value. The present tool predicts docking result by obtaining energy value for each ligand. Finally, all ten ligands were analysed to detect binding position and energy value. The resultant structural complexes of the individual ligand/receptor binding were finally recorded to determine some specific contacts between the atoms of

the test compounds and amino acids of the DNA gyrase B protein. In the post-screening analysis, hydrogen bonding value, van der Waal and electrostatic forces were obtained to know the efficiency of protein-ligand interaction clustering.

## RESULTS AND DISCUSSION

Present computational prediction (molecular docking) indicates that favourable binding energy was observed in 5-hydroxy-7,8,2',5'-tetramethoxy-flavone (-121.862 Kcal/mol) when compared to Ciprofloxacin (-106.621 Kcal/mol), among other flavonoids of *A. paniculata*. In case of other flavonoids, the binding energy values (Kcal/mol) of all 9 flavonoids such as Andrographolide (-116.083), Andrographidine C (-116.083), 7-O-methylwogonin (-110.152), 5,7,3',4'-tetrahydroxyflavone (-109.642), 5,7,4'-trihydroxyflavone (-102.227), 7-O-methyl dihydrowogonin (-95.3081), Neoandrographolide (-91.8332) and 14-Deoxyandrographolide (-84.1036) respectively (Table 1).

In case of interaction study, the close contact residues such as ILE51, ILE175, THR173, ILE86, ASP81, GLY85, ARG84 and GLU58 and one hydrogen bond contact residue as ASN54 in chain A were obtained for 5-hydroxy-7,8,2',5'-tetramethoxy-flavone while the contact residues such as PRO87, ILE86, THR173, GLY, 85, ASP84, ARG84, GLU58 and ASN54 in chain A without hydrogen bond contact were observed for Ciprofloxacin (Fig 2). On the other hand, the eight studied flavonoids were also observed the binding

interaction with contact residues and hydrogen bonding (Fig 3). For Andrographolide and Andrographidine C, the contact residues such as ILE51, ASN54, GLU58, ARG84, ARG144, GLY85, ASP81, ILE86 and THR173 and four hydrogen bond contacts, one with SER55 and others with inhibitory molecule (B471) in chain A were observed. For 7-O-methylwogonin, ILE51, ASN54, GLU58, ARG84, THR173, ILE86, PRO87, ILE175 and ILE102 as contact residues and two hydrogen bond contacts, one with GLY85 and other with inhibitory molecule (B471) in chain A were observed. For 5,7,3',4'-tetrahydroxyflavone, the contact residues such as ILE51, GLU58, ARG84, ASP81, GLY85, ILE86, THR173 and ILE175 and one hydrogen bond contact with ASN54 in chain A were observed. For 5,7,4'-trihydroxyflavone, the contact residues such as ILE51, ILE175, THR173, ASP81, ILE86, GLY85, GLU58 and hydrogen bonding with ASN54 in chain A were obtained. For 7-O-methyl dihydrowogonin, the contact residues such as ASP87, GLU58, ASP81, THR173 and ILE175 and one hydrogen bond contact with ASN54 in chain A were observed. For 14-Deoxyandrographolide, the contact residues such as ASP57, SER55, GLU58, ARG84, THR173, ILE86, PRO87, ILE175 and ASN54 and one hydrogen bond contact with inhibitory molecule (B471) in chain A were observed. For Neoandrographolide, the contact residues such as ASN54, SER55, GLU58, THR173, GLY85, ILE86, ILE102, SER128 and hydrogen bonding with inhibitory molecule in chain A were obtained.

**Table 1: Interaction profiles of selected ligands of *A. paniculata* and known antibiotic after docking against bacterial DNA gyrase B receptor.**

Sl. No.	Ligands	Binding energy (Kcal/mol)	vander Waals value	Hydrogen bonding value	Electrostatic Bonding value
<b>Phytochemicals</b>					
1.	5-hydroxy-7,8,2',5'-tetramethoxy-flavone	-121.862	-97.2159	-24.6463	0
2.	Andrographolide	-116.083	-98.4045	-17.6786	0
3.	Andrographidine C	-116.039	-98.1866	-17.8523	0
4.	7-O-methylwogonin	-110.152	-89.9666	-20.1858	0
5.	5,7,3',4'-tetrahydroxyflavone	-109.642	-81.6725	-27.9695	0
6.	5,7,4'-trihydroxyflavone	-102.227	-85.8295	-16.3975	0
7.	7-O-methyl dihydrowogonin	-95.3081	-81.6637	-13.6443	0
8.	Neoandrographolide	-91.8332	-70.3942	-21.439	0
9.	14-Deoxyandrographolide	-84.1036	-59.3194	-24.7842	0
<b>Synthetic chemical</b>					
10.	Ciprofloxacin	-106.621	-92.681	-15.9616	2.02182

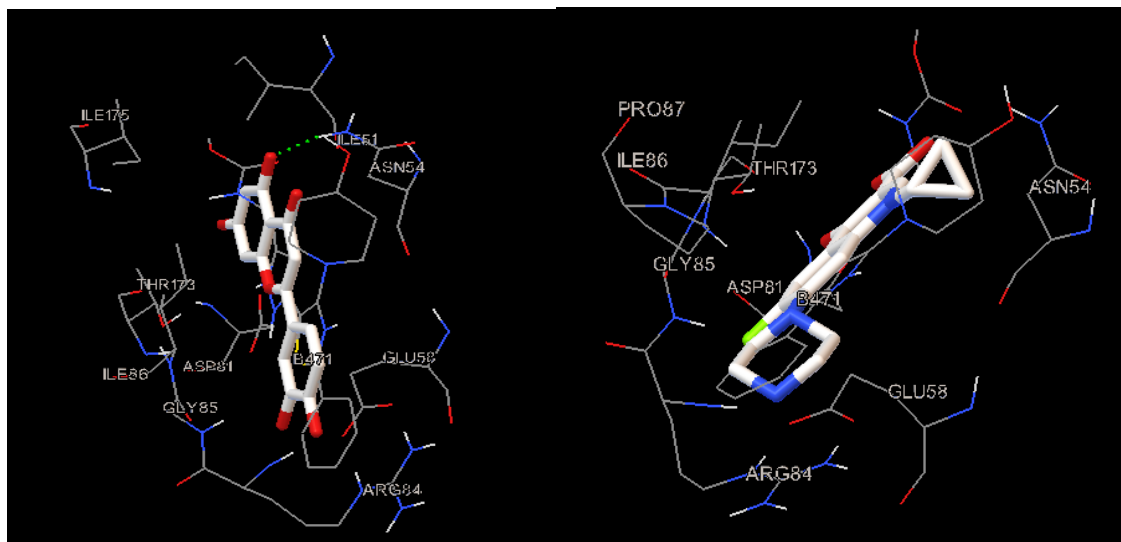
It was reported in earlier studies that active site residues are found in DNA gyrase B such as GLU50, ASN54, GLU58 and THR173 present in the ATP binding pocket, which are involved in ATPase activity (Gross et al., 2003; Jagadeesan et al., 2015)32,33. In present prediction with the flavonoid phytoligand (5-hydroxy-7,8,2',5'-tetramethoxy-flavone), and synthetic ligand (Ciprofloxacin) both were found binding to the active site in chain A of bacterial DNA gyrase B (PDB ID: 3G7B).

But phytoligand 5-hydroxy-7,8,2',5'-tetramethoxy-flavone showed highest binding energy among other studied flavonoids, which may be due to competitive inhibition. In another research work, it was observed that phytoligand Epigallocatechin-3-gallate higher activity to inhibited bacterial DNA gyrase B by interaction with its ATP binding site and these catechins are containing in green tea (Gradišar et al., 2007)34. According to Barik and Talukdar (2018),35 phytoligand as flavonoid

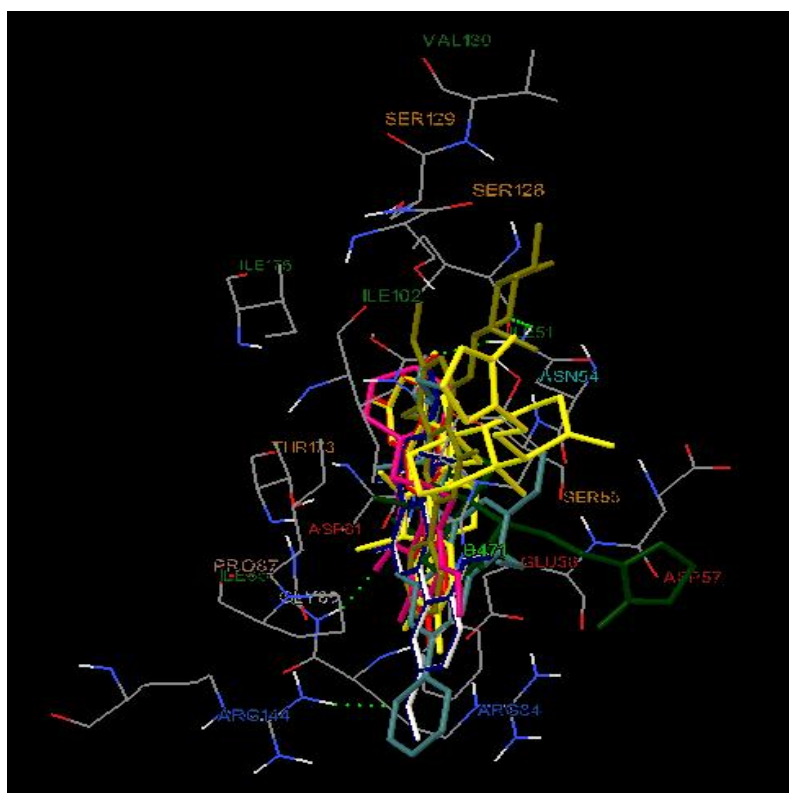
Epigallocatechin-3-gallate showed highest binding energy and binding opposite side of the active site due to non-competitive inhibition. Although, researchers documented in experimental study that the crude extract of *A. paniculata* is well-known antibacterial agent but the function of exact phytochemical in antimicrobial activity is unknown. The selection of phytochemicals of *A. paniculata* in the present computational prediction on the basis of literatures supported secondary metabolites

especially flavonoids have already been investigated as antibacterial natural compound.

The interaction profile of post-screening analysis for protein-ligand complexes showed for receptor-ligand binding efficiency and hierarchical tree representation for the compound similarities as obtained through the present tool (Fig 4).

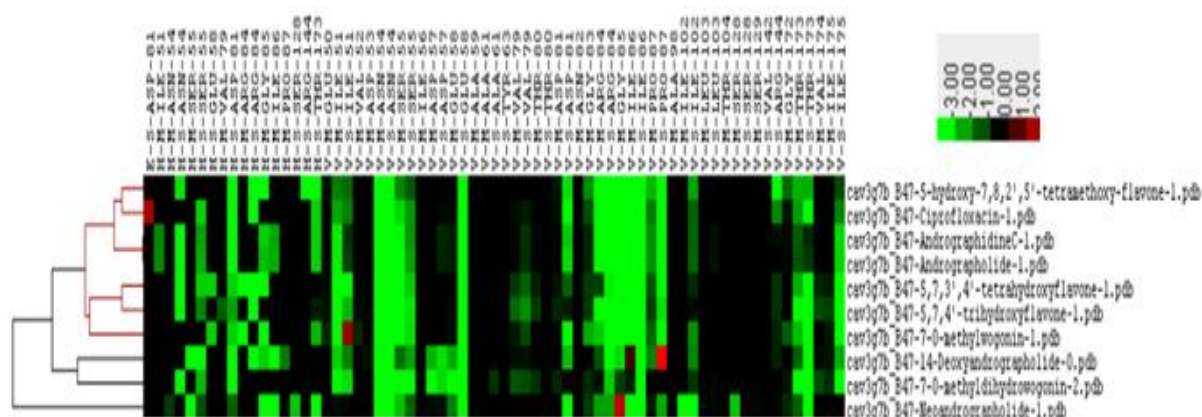


**Fig. 2: 5-hydroxy-7,8,2',5'-tetramethoxy-flavone and Ciprofloxacin docking interactions.**



**Fig. 3: Docking interactions of Andrographolide, Andrographidine C, 7-O-methylwogonin, 5,7,3',4'-tetrahydroxyflavone, 5,7,4'-trihydroxyflavone, 7-O-methyldihydrowogonin, Neoandrographolide and 14-Deoxyandrographolide.**





**Fig. 4: Hierarchical tree representation for the compound similarities using receptor-ligands interaction (Interactions as E = electrostatic; H = hydrogen-bonding and V = vander Waals; M and S = main chain and side chain of the interacting residue) along with colour code bar value.**

It was obtained by the present tool a combinatorial cluster analysis and visualize compound candidates generated by virtual screening (VS) through hierarchical tree representation. Basically, clustering steps help to know interaction specific information between target and lead compound(s). The hydrogen bonding value, van der Waal and electrostatic forces are important in receptor-ligand binding increasing the efficiency of protein-ligand interaction clustering. According to Clinciu et al. (2010)36, the combination of an optimized docking tool and two clustering stages for the scope of selecting ideal representatives revealed promising results.

It was established that the vdW interactions are coloured in green when the energy is less than -4. The hydrogen bonding and electrostatic interactions are coloured in green if the energy is  $\leq 24$ . M. Hydrogen bonding and van der Waals interactions are the major contributors to compounds recognition in the target-receptor (Kandeeland Kitade Y, 2013)37.

## CONCLUSION

It is concluded from the computational prediction by molecular docking approach the flavonoid phyto ligand 5-hydroxy-7,8,2',5'-tetramethoxy-flavone can be alternative of synthetic antibacterial drug as per binding energy value and interaction of competitive inhibition. According to Zhang et al., (2016)38 plant derived natural chemical can be suitable as lead compound to prevent synthetic drug resistance of bacteria. This present *in silico* study is suggested to validate further toxicological and pharmacological assay with this flavonoid after isolation from medicinal plant (*A. paniculata*) on different pathogenic bacteria caused plant pathogenicity, which can be prevented bacterial infection on plants without resistance and environmental hazards.

## ACKNOWLEDGEMENT

The authors convey thanks to the developers of present software used in the predictive study, data bank for protein and phytochemicals.

## Conflict of interest

None

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